**Niche partitioning among and within sympatric tropical seabirds revealed by stable isotope analysis**

Hillary S. Young1,*, Douglas J. McCauley1, Rodolfo Dirzo1, Robert B. Dunbar2, Scott A. Shaffer3

1Department of Biology and 2Environmental Earth System Science, Stanford University, Stanford, California 94305, USA
3Department of Biological Sciences, San José State University, San José, California 95192, USA

ABSTRACT: The low productivity and unpredictable nature of resources in tropical waters would appear to make resource partitioning among predators difficult. Yet, stable isotope data from the present study suggest that substantial resource partitioning occurs among tropical seabird communities, both between and within species. In the present study, we compared \( \delta^{13}C \) and \( \delta^{15}N \) levels among 8 sympatric tropical seabird species. For a subset of these species, we also examined isotopic levels across years, breeding periods, and sexes. When breeding and non-breeding periods were both considered, we found that all species occupied distinct isotopic niches across at least one time period. Resource partitioning by age (within the same year) and year studied (for birds of the same age), within species was observed in all the species we examined. However, niche separation by sex appeared to be relatively uncommon even in sexually dimorphic species. As a group, seabirds were isotopically distinct from other marine predators. There was a strong correlation between both body mass and wing loading ratios and foraging area, as evaluated using \( \delta^{13}C \) levels. Cumulatively, the isotopic patterns observed are consistent with diet-based surveys of resource partitioning during the breeding period. Importantly, these data provide additional evidence to suggest that substantial niche partitioning among tropical seabird species persists even during non-breeding periods in open ocean environments, when direct diet-based surveys are difficult to conduct.

KEY WORDS: Resource partitioning · Foraging ecology · \( \delta^{13}C \) · \( \delta^{15}N \) · Marine predators

INTRODUCTION

Tropical open ocean environments generally have lower productivity and lower structural complexity, and thus patchier and more unpredictable distributions of prey than temperate oceans (Longhurst & Pauly 1987, Ballance et al. 1997). High trophic level marine predators thus face many challenges locating prey and have likely evolved behavioral and morphological adaptations that facilitate energetically efficient travel and foraging behavior (Bertrand et al. 2002, Weimerskirch et al. 2004). Despite these constraints, diverse communities of marine predators, particularly seabirds, exist, leading to questions about the degree, nature, and mechanisms of resource partitioning in these relatively unpredictable and nutrient-limited environments.

Resource partitioning in oceanic environments has been well documented for temperate and polar seabird species (Ainley et al. 1992, 1994), but remains an area of much inquiry for tropical species, particularly in open ocean environments (Harrison & Seki 1987, Catry et al. 2009). Tropical seabird diets are more diverse than their temperate and polar counterparts, and there can be a high degree of overlap in diets and foraging areas (Ballance et al. 1997, Catry et al. 2009). Most tropical species are also limited to foraging within the first several meters of the sea surface, and they often rely on subsurface predators to drive food to the surface, further increasing the patchy nature of food resources (Ballance et al. 1997). These observations have led to questions about the degree to which tropical seabird species are able to partition resources.

*Email: hyoung@stanford.edu

© Inter-Research 2010 · www.int-res.com
While there have been several comprehensive extensive studies of the diets of seabird communities (using stomach contents) in tropical environments, these have generally been constrained to the breeding season (Ashmole & Ashmole 1967, Harrison et al. 1983, Catry et al. 2009). The only study to comprehensively explore the diet contents of non-breeding and breeding tropical seabird communities found higher degrees of resource partitioning than found in a similar study of polar seabirds (Spear et al. 2007). In that study, Spear et al. (2007) found evidence of partitioning by species, sex, age, foraging strategy, and body size. There was even a high degree of prey separation among species foraging in the same flocks. However, Spear et al. (2007) found incomplete partitioning among species, and several of the species were statistically indistinguishable from one another.

Despite the many strengths of at-sea surveys of seabird diet contents and the detail of data resolution achieved, this methodology has some inherent limitations for answering questions about resource partitioning. Since seabirds are typically sampled lethally, the stomach contents from each animal contribute only a single data point in space and time, and it is thus not possible to examine changes in individual foraging behavior across space and time. Also, it is not possible to link foraging behavior to reproductive status using these methods, although it is likely that reproductive status influences foraging behavior (Schaffer et al. 2003). Additionally, collections across long time periods may compare foraging across heterogeneous time periods.

Stable isotope analysis (SIA) has been shown to provide an excellent complementary tool for understanding resource partitioning, both spatially and by trophic level (Bearhop et al. 2002). Stable isotopes of carbon and nitrogen in seabird tissue are derived from the isotopic values of their prey and give insight into the origin and type of prey consumed (although they do not provide detailed information on prey identity or capture method, which is available from dietary analysis or direct observation). Stable carbon isotopes show evidence of an increasing enrichment in inshore as compared to offshore feeding animals in both tropical and temperate areas (Cherel et al. 2008). For tropical surface-feeding seabirds, benthic to surface gradients in carbon isotopes are likely not directly relevant. However, in comparing seabirds to other marine predators, the enriched carbon values found in benthic habitats are also an important consideration. Stable nitrogen isotopes increase in a predictable matter with each trophic step, and thus indicate trophic position of a consumer (Vanderklift & Ponsard 2003). There do not seem to be strong species-specific fractionation rates among fish-eating seabirds (Cherel et al. 2005a), so differences across species within a system are likely due to diet differences. Comparisons across systems must be undertaken with more caution; clear and strong gradients across latitudes exist both for carbon (>10‰, with less negative values in the tropics) and for nitrogen (>6‰, with lower values in the tropics; Graham et al. 2010). Since different tissues (e.g. feathers, blood) integrate these stable isotopes over different periods of time, a single bird can provide integrated information on diet over a period of time ranging from days to years, depending on the tissues analyzed (Dalerum & Angerbjörn 2005).

SIA has yielded important insights into subtle changes in seabird foraging ecology based on reproductive stage (e.g. Awkerman et al. 2007), age (e.g. Forero et al. 2002), colony location (e.g. Jaquemet et al. 2008), sex (e.g. Bearhop et al. 2006), migratory patterns (e.g. Cherel et al. 2000), as well as other factors (e.g. Cherel et al. 2005b, Phillips et al. 2009). Since SIA is a robust, non-lethal method to resolve spatial and trophic separation among species, it is a useful tool to examine resource partitioning among seabirds in tropical environments, particularly for those species that are too small for current electronic tracking methods. A few studies have used SIA to examine resource partitioning on a community scale in the tropics (Catry et al. 2008, Cherel et al. 2008, Kojadinovic et al. 2008). Those studies, all conducted in the Southern Indian Ocean, showed relatively small variation in stable isotopes across species (<2‰ variation across species, within a tissue type). However, they reached different conclusions, both in the degree of resource partitioning and the usefulness of isotopes as a tool for identifying foraging patterns in the tropics.

Our study site, Palmyra Atoll (equatorial Pacific), is quite different from the other tropical study sites where SIA has been conducted, in that Palmyra is far (1000s of km) from any continental shelf and has lower heterogeneity in oceanic conditions (productivity, sea surface temperature, bathymetry; Weimerskirch et al. 2005, Young et al. 2010). This site may thus more generally typify the open ocean conditions experienced by many tropical seabirds, particularly during the non-breeding period. It also has a remarkably intact surrounding marine community, with naturally high biomass of high trophic level predators (Stevenson et al. 2007). This, combined with the high number of co-occurring nesting species examined in the present study, creates an ideal situation to examine niche partitioning within and among these marine predators.

The aims of the present study were to (1) determine if there is isotopic evidence for significant resource partitioning among tropical seabird species living in an open and relatively intact ocean environment; (2) compare patterns of isotopic partitioning in blood and
feathers to evaluate differences in partitioning among species in breeding (blood) and non-breeding (feather) periods; (3) examine isotopic evidence of resource partitioning within species across different age classes and sexes; and (4) compare seabird resource use to that of other marine predators (predatory fish and invertebrates), with whom seabirds must also compete for resources. We compared our stable isotope results to data from conventional stomach content analyses, at-sea surveys, and known natural history. These comparisons included multiple congeneric species, and species with similar foraging strategies, where fine-scale niche partitioning might be particularly important for mitigating competitive interactions.

**MATERIALS AND METHODS**

**Study site and species.** Our research was conducted at Palmyra Atoll National Wildlife Refuge (5.867°N, 162.067°W). Palmyra Atoll is a tropical atoll located in the Line Island chain in the central Pacific Ocean. It is situated on the boundary of the intertropical convergence zone (Longhurst & Pauly 1987). The surrounding waters are generally low in productivity, warm, and deep (Young et al. 2010). The seabird community at Palmyra consists of 10 breeding species in 2 orders: (1) Charadriiformes: sooty terns *Sterna fuscata* (~125 000–220 000 pairs), white terns *Gygis alba* (~250 pairs), brown noddy *Sula sula* (~250 pairs), red-footed boobies *S. dactylatra* (~35 pairs), red-tailed tropicbird *Phaethon rubricauda* (~150 pairs), and white-tailed tropicbirds *P. lepturus* (~10 pairs) (Fefer 1987, H. S. Young unpubl. data). At Palmyra these birds breed asynchronously throughout the year.

**Sample collection.** We collected feather samples from 9 of the 10 species that breed at the atoll (all except white-tailed tropicbirds; Table 1). Samples were collected from breeding adults (either incubating or chick rearing), with the exception of samples from white terns, where reproductive status was unknown. All samples were collected in July 2009, except for sooty terns, which did not breed in summer 2009; for this species, we thus used feathers collected in July 2008. For red-footed boobies, additional samples were collected from breeding adults in 2 previous years (July 2007, *n* = 15; and July 2008, *n* = 25) for interannual comparisons. To examine resource differentiation between parents and offspring (e.g. via selective feeding), for 5 species we also analyzed feathers collected from unfledged chicks in 2009 (black noddy, *n* = 8; red-footed booby, *n* = 10; red-tailed tropicbird, *n* = 15; brown booby, *n* = 10). For red-footed boobies only, we also compared immature birds (fledged, but not yet in adult plumage; *n* = 10) to adults and chicks. Since feathers of adults are formed in the non-breeding period, while those of chicks and juveniles are formed during the breeding period, differences observed may also be due to foraging differences across these time periods. Four sexually dimorphic species were examined for sexual differences in resource partitioning (red-footed booby, *n* = 6 per sex; brown booby, *n* = 8 per sex; masked booby, *n* = 7 per sex; great frigatebird, *n* = 7 female and 11 male).

<table>
<thead>
<tr>
<th>Species</th>
<th>n</th>
<th>$\delta^{15}$N</th>
<th>$\delta^{13}$C</th>
<th>C:N</th>
</tr>
</thead>
<tbody>
<tr>
<td>Feathers: seabirds</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Black noddy</td>
<td>18</td>
<td>14.79 ± 1.90$^{AB}$</td>
<td>-16.87 ± 0.32$^{A}$</td>
<td>3.34 ± 0.07</td>
</tr>
<tr>
<td>Brown noddy</td>
<td>3</td>
<td>16.35 ± 0.31</td>
<td>-16.12 ± 0.25</td>
<td>3.37 ± 0.01</td>
</tr>
<tr>
<td>Sooty tern</td>
<td>21</td>
<td>14.7 ± 2.80$^{AB}$</td>
<td>-16.30 ± 0.28$^{B}$</td>
<td>3.23 ± 0.03</td>
</tr>
<tr>
<td>White tern</td>
<td>20</td>
<td>16.20 ± 1.69$^{A}$</td>
<td>-15.91 ± 0.29$^{C}$</td>
<td>3.16 ± 0.04</td>
</tr>
<tr>
<td>Brown booby</td>
<td>21</td>
<td>14.08 ± 1.39$^{B}$</td>
<td>-15.43 ± 0.31$^{D,E}$</td>
<td>3.18 ± 0.03</td>
</tr>
<tr>
<td>Masked booby</td>
<td>15</td>
<td>14.17 ± 0.98$^{B}$</td>
<td>-15.22 ± 0.24$^{E}$</td>
<td>3.18 ± 0.03</td>
</tr>
<tr>
<td>Red-footed booby</td>
<td>12</td>
<td>13.85 ± 1.25$^{B}$</td>
<td>-15.73 ± 0.34$^{C,D}$</td>
<td>3.22 ± 0.03</td>
</tr>
<tr>
<td>Great frigatebird</td>
<td>19</td>
<td>16.03 ± 1.09$^{A}$</td>
<td>-16.26 ± 0.20$^{B}$</td>
<td>3.27 ± 0.03</td>
</tr>
<tr>
<td>Red-tailed tropicbird</td>
<td>26</td>
<td>15.35 ± 1.70$^{AB}$</td>
<td>-15.61 ± 0.27$^{D}$</td>
<td>3.14 ± 0.03</td>
</tr>
<tr>
<td>Blood: seabirds</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sooty tern</td>
<td>17</td>
<td>14.29 ± 1.52$^{A}$</td>
<td>-18.02 ± 0.13$^{A}$</td>
<td>3.33 ± 0.11</td>
</tr>
<tr>
<td>Brown booby</td>
<td>10</td>
<td>15.67 ± 0.72$^{B}$</td>
<td>-17.20 ± 0.12$^{B}$</td>
<td>3.30 ± 0.03</td>
</tr>
<tr>
<td>Masked booby</td>
<td>9</td>
<td>14.21 ± 0.61$^{A}$</td>
<td>-17.15 ± 0.18$^{C}$</td>
<td>3.40 ± 0.06</td>
</tr>
<tr>
<td>Red-footed booby</td>
<td>11</td>
<td>13.79 ± 0.44$^{A}$</td>
<td>-17.42 ± 0.09$^{D}$</td>
<td>3.31 ± 0.04</td>
</tr>
<tr>
<td>Great frigatebird</td>
<td>5</td>
<td>16.07 ± 0.72$^{B}$</td>
<td>-17.20 ± 0.12$^{C,D}$</td>
<td>3.41 ± 0.03</td>
</tr>
<tr>
<td>Muscle: other predators</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acanthocybium solandri</td>
<td>15</td>
<td>14.53 ± 3.15</td>
<td>-16.21 ± 1.24</td>
<td>3.15 ± 0.06</td>
</tr>
<tr>
<td>Thunnus albacores</td>
<td>25</td>
<td>14.63 ± 0.96</td>
<td>-16.68 ± 0.36</td>
<td>3.16 ± 0.15</td>
</tr>
<tr>
<td>Sthenoteuthis spp.</td>
<td>11</td>
<td>12.82 ± 1.02</td>
<td>-17.62 ± 0.27</td>
<td>3.18 ± 0.05</td>
</tr>
</tbody>
</table>

Table 1. Stable isotope ratios of carbon and nitrogen and mass ratio of C:N in feathers and blood of seabirds, and muscle tissue of other pelagic predators. For seabirds, superscript letters denote significant differences between species in post hoc analyses; values not sharing the same letter are significantly different (comparisons are within tissue type). Brown noddies were not considered in post hoc comparisons due to small sample size. Values are mean ± SD.
Field metabolic rate (kJ d⁻¹) was estimated from allometric equations derived for Pelicaniformes and Charadriiformes (Shaffer in press).

To compare seabirds to other pelagic predators, muscle tissue from pelagic fish predators, wahoo Acanthocybium solandri (n = 15) and yellowfin tuna Thunnus albacares (n = 25), was collected from animals captured from within 3 km of the reef immediately surrounding the atoll during July to August of multiple years (2007 to 2009). Flying squid Sthenoteuthis spp. (n = 11) were captured approximately 700 km from the atoll (May 2007). Comparisons among seabirds and other marine predators only included the 5 seabird species for which blood data were available (as feathers are likely not directly comparable to muscle tissue; Hobson & Clark 1992).

**Sample preparation and isotope analyses.** Feathers were washed in deionised water, dried at 60 °C for storage, and subsequently cut into fine pieces for analysis. Blood and muscle samples (from seabirds and other marine predators) were all preserved frozen at –80 °C. They were then freeze dried and ground to a fine powder. We did not extract lipid from any tissues, as C:N ratios were always less than 4.0, and usually less than 3.5, suggesting lipid levels were low across all samples (Post et al. 2007).

Stable isotopic ratios of C and N were analyzed at the Stanford Stable Isotope Biogeochemistry Laboratory (SIBL) using a Thermo Finnigan Delta-Plus XP IRMS coupled to a Carlo Erba NA 1500 Series 2 elemental analyzer. Replicate laboratory standards of graphite NIST RM 8541 (USGS 24), and ammonium sulfate NIST RM 8547 (IAEA N1) internal to each run showed an analytical error of <0.2 ‰ for both C and N.

**Isotope interpretations.** The interpretation of carbon isotope values presented in this paper is based on the assumption that the established inshore to offshore gradient of δ¹³C (Cherel & Hobson 2007, Graham 2009) is likely the primary driver for changes in stable carbon isotopes of the seabirds measured in the present study. Although Palmyra Atoll has a limited reef area, a δ¹³C gradient exists between reef fish and more pelagic fish (D. J. McCauley et al. unpubl. data). Changes in carbon isotopes due to trophic level differentiation are presumed to be generally small in seabirds, and given the limited range of δ¹³C observed in the present study, trophic variation is not likely to be an important explanatory factor of δ¹³C here (Kelly 2000). Variation in δ¹⁵N within a tissue type is likely primarily due to differences in trophic position.

We further assumed that there is no size or age specific fractionation rates of either carbon (ΔC) or nitrogen (ΔN) isotopes within seabirds (Cherel et al. 2005a). We did not directly compare δ¹⁵N or δ¹³C across tissue types, given different isotopic signatures and fractionation rates of these tissues (Cherel et al. 2005a). The period of isotopic integration in blood was assumed to be days to weeks, such that blood taken from a breeding bird is assumed to give diet information on the breeding period (Hobson & Clark 1992). Since feathers are usually molted after reproduction, and are inert thereafter, feather samples were assumed to represent the composition of the diet during the non-breeding period (Bearhop et al. 2002).

**Statistical analyses.** To examine differences in resource partitioning among species, we used multivariate analysis of variance (MANOVA), with subsequent univariate ANOVA tests of differences in δ¹³C and δ¹⁵N, with post hoc Tukey HSD analyses. Due to the small sample size of brown noddies, they were not considered in post hoc analyses. Statistical analyses were performed in JMP 8 (SAS Institute). When necessary to meet assumptions of normality and homoscedasticity, data were transformed using Box-Cox transformation. All figures and tables depict untransformed data. All data are reported as untransformed means ± 1 SD.

**RESULTS**

**Resource partitioning among species in breeding and non-breeding periods**

The 9 species of adult seabirds showed significant overall isotopic segregation in feather samples (MANOVA, Wilks’ lambda, $F_{16,288} = 12.49, p < 0.0001$; Fig. 1A). Univariate tests of feathers across species also showed significant differences in both δ¹³C (ANOVA, $F_{3,145} = 30.41, p < 0.0001$) and δ¹⁵N (ANOVA, $F_{3,145} = 5.20, p < 0.0001$). Results from post hoc Tukey pairwise comparisons showed significant differences in δ¹³C among most species, although not between every species pair (Table 1). For δ¹⁵N, the 3 booby species were all significantly different from the white tern and the great frigatebird; there were no other significant differences (Table 1).

The 5 species sampled for blood also showed significant overall isotopic segregation (MANOVA, Wilks’ lambda, $F_{3,90} = 36.63, p < 0.0001$; Fig. 1B). Univariate tests showed significant differences both for δ¹³C (ANOVA, $F_{4,46} = 129.10, p < 0.0001$) and for δ¹⁵N (ANOVA, $F_{4,46} = 30.41, p < 0.0001$). In post hoc analyses, all seabird species had distinct values for δ¹³C, except great frigatebirds, which were indistinguishable from either masked or red-footed boobies (Table 1). For δ¹⁵N, great frigatebirds and brown boobies were statistically different from the other 3 species (Table 1).

All seabirds differed from one another in at least 1 of the 2 stable isotopes measured in blood or feathers. The total range spanned in mean values among spe-
Species was 1.2‰ (in feathers) and 1.8‰ (in blood) for \( \delta^{13}C \), and 2.5‰ (in feathers) and 2.3‰ (in blood) for \( \delta^{15}N \). The pattern of trophic partitioning among species changed somewhat between blood and feathers, with great frigatebirds having a relatively lower \( \delta^{13}C \) in feathers compared to blood, and brown boobies a relatively higher \( \delta^{13}C \) in feathers compared to blood.

Comparison of mean \( \delta^{13}C \) by mean body mass per species yielded a significant positive relationship (\( R^2 = 0.67, p < 0.01 \)), with higher \( \delta^{13}C \) enrichment in larger birds (Fig. 2). The relationship improved further when \( \delta^{13}C \) was compared to wing loading (\( R^2 = 0.82, p < 0.01 \)), where birds with lower wing loading were more enriched with \( \delta^{13}C \). There was also a strong positive relationship between estimated field metabolic rate and \( \delta^{13}C \) (\( R^2 = 0.74, p < 0.01 \)). There were no significant relationships between any of the above variables and \( \delta^{15}N \) values.

Of the 5 species for which adult and chick feathers were compared, 4 showed significantly higher \( \delta^{15}N \) values in chicks than in adults (black noddy, \( t = 2.47, p = 0.02 \); red-footed booby \( t = 2.92, p = 0.01 \); red-tailed tropicbird, \( t = 5.63, p < 0.0001 \); brown booby \( t = 2.71, p = 0.01 \)). For red-footed boobies, when juvenile birds were also considered there were similarly significant effects (MANOVA, Wilks’ lambda, \( F_{4,44} = 3.21, p = 0.02 \)), with chicks having slightly higher \( \delta^{15}N \) values (\( F_{2,23} = 5.49, p = 0.01 \)) than either adult or juvenile birds (Fig. 3). Great frigatebirds were the only species to show a marginally significant increase in \( \delta^{15}N \) (\( t = 1.82, p = 0.08 \); Fig. 4). Only 1 species, red-tailed tropicbirds, showed significant differences in \( \delta^{13}C \) by age, with adult birds having lower \( \delta^{13}C \) than chicks (\( t = 5.63, p < 0.0001 \)).

Among the 4 species analyzed for effects of sex on isotopes, significant differences were found for only 1 species, the brown booby. Male brown boobies had lower \( \delta^{13}C \) (\( t = 4.23, p < 0.001 \)) and lower \( \delta^{15}N \) (\( t = 2.40, p = 0.03 \)) than females.

There was a slight but significant interannual difference in \( \delta^{13}C \) of feathers for breeding adult red-footed boobies (ANOVA, \( F_{4,126} = 4.10, p < 0.01 \)), where boobies in 2007 had slightly higher \( \delta^{13}C \) values than birds in either 2008 or 2009 (ANOVA, \( F_{2,50} = 3.61, p = 0.03 \); Fig. 3). No such trend was detected in \( \delta^{15}N \).

**Comparisons of seabirds to other marine predators**

As a group, seabirds were isotopically distinct from other large pelagic predators (MANOVA, Wilks’
Lambda, $F_{6,298} = 23.26, p < 0.0001$; Fig. 5) with large predatory fish (wahoo, yellowfin tuna) having higher $\delta^{13}C$ than seabirds or the large predatory Sthenoteuthis squid ($F_{6,149} = 64.93, p < 0.0001$). Sthenoteuthis spp. had lower $\delta^{15}N$ than seabirds or large predatory fish ($F_{6,149} = 15.87, p < 0.0001$). On a species by species basis there was some overlap, with brown boobies being statistically indistinguishable from yellowfin tuna, and red-footed boobies being statistically indistinguishable from Sthenoteuthis spp. (Fig. 5).

**DISCUSSION**

There were significant differences in either carbon or nitrogen isotopes among all species examined in either breeding or non-breeding periods. This suggests that all the sympatric seabird species examined may be occupying distinct ecological niches, when both breeding and non-breeding periods are considered. The degree of partitioning observed in the non-breeding period was more complete than the partitioning detailed by at-sea surveys and diet analyses (Surman & Wooller 2003, Spear et al. 2007).

**Resource partitioning by species in the non-breeding period**

The patterns observed in $\delta^{13}C$ in the non-breeding period (feathers) were generally consistent with data from tracking and at-sea surveys. Sooty terns and great frigatebirds have the most pelagic signal, followed sequentially by white terns, red-footed boobies, red-tailed tropicbirds, brown boobies and masked boobies (Bal lance et al. 1997, Jaquemet et al. 2005). One surprising result based on carbon isotope levels was the highly pelagic signal of black noddies (i.e. values were more negative than that of any other species). This species is generally considered to be an opportunistic nearshore feeder that is often seen foraging near jacks and within lagoons (Ashmole 1968, Seki & Harrison 1989). Yet values of
carbon isotope levels in lagoon waters at Palmyra are particularly elevated, and black noddy isotope levels do not resemble those of reef jacks at Palmyra (D. J. McCauley et al. unpubl. data). Studies of procellariiforms have found that relatively negative δ13C values can derive from the assimilation of carbon from dietary lipids (Thompson et al. 2000). It is possible that black noddies have differential abilities to incorporate dietary lipids than other species studied, although this seems unlikely given the lack of similarly negative δ13C in brown noddies. Given the small total range of δ13C observed, controlled measurements of species-specific fractionation rates would be helpful to confirm that species-specific fractionation rates do not drive these patterns (Becker et al. 2007). Likewise, while δ13C maps available for the equatorial Pacific Ocean do not suggest high variation in δ13C around this region, that is based on limited sampling near Palmyra (Graham 2009); thus, better fine-scale isotopic sampling of oceans in this region would improve the accuracy of our interpretations.

Species-level changes in δ13C were highly correlated to body mass, with smaller birds exhibiting a more pelagic signature than larger species. Since body mass co-varies with wing loading and with metabolic rate, it is possible that either or both of these variables may partially drive these patterns. However, since laboratory manipulations of metabolic rate in birds via exercise and cold show no effect of metabolic rate on ΔC, it seems unlikely that differential metabolic rates drive these patterns (Bauchinger et al. 2010).

Nitrogen isotopes from non-breeding periods (feathers) showed that great frigatebirds had elevated δ15N levels over many species, including all the boobies (consistent with Cherel et al. 2008). When at the colony, these birds also often consume chicks of various smaller species and steal partially regurgitated food from other birds, potentially explaining this variation (Schreiber & Hensley 1976). As found in the Seychelles (Catry et al. 2008), white terns also showed high δ15N levels. While Catry et al. (2008) suggested that this might be reason to discount results of δ15N, the consistency across studies perhaps merits further consideration for biologically valid explanations. It is possible that these high levels may be due to the large portion of its diet (> 40%) composed of small, predatory Scombridae (Euthynnus sp.; Spear et al. 2007). This predatory species might be higher in δ15N than flyingfish and squid, which dominate the diet of several other species. Direct measurements of δ15N of these prey would be necessary to resolve this result. The 3 booby species, which feed primarily on flyingfish and squid (Schreiber & Hensley 1976, Spear et al. 2007) showed particularly low δ15N values. Controlled studies of species-specific fractionation patterns would help clarify if physiological factors or trophic differences are influencing δ15N patterns (Becker et al. 2007).

Resource partitioning by species in the breeding period

Examination of blood samples, representing the breeding interval, also showed distinct niches for each of the species examined. There were small changes in foraging areas in non-breeding as opposed to breeding periods, with great frigatebirds and brown boobies appearing comparatively less pelagic than during the non-breeding interval. For the 2 species for which tracking data were available at Palmyra (red-footed and masked booby), the δ13C results were highly consistent with tracking results (with red-footed boobies foraging slightly farther off shore than masked boobies during the breeding period), even though the differences in foraging distances were not extremely large (Young et al. 2010).

The relative positions of δ13C and δ15N in both breeding and non-breeding periods observed at Palmyra were highly consistent with those observed on Europa Island and in the Seychelles (Catry et al. 2008, Cherel et al. 2008), suggesting that niche partitioning is consistent in very different parts of the species’ ranges. The absolute values of δ13C were also consistent in both studies in the non-breeding interval, but slightly depleted at Palmyra during the breeding interval, perhaps due to the more oceanic location of Palmyra itself, as open ocean conditions likely lead to lower δ13C values (Graham et al. 2010). In contrast, absolute values of δ15N observed at Palmyra were greatly elevated (by about 2‰) over those obtained at Europa in both breeding and non-breeding periods. This suggests either a higher baseline of nitrogen at Palmyra than at Europa, or a higher overall trophic position of birds at Palmyra. Similar results were seen in a comparison of the Seychelles to the Mozambique Channel (Jaquemet et al. 2008). This is consistent with latitudinal variations of δ15N in isotopic cartography, suggesting that while it is possible to compare relative δ15N positions across studies, to compare absolute values of δ15N, good knowledge of baseline δ15N is needed (Graham et al. 2010).

Partitioning within species

A comparison of feathers from chicks and adults showed elevated levels of δ15N in chicks across all 5 species examined. This differed from the pattern seen for great frigatebirds on Europa Island, but is consistent with results for sooty terns at the same site (Cherel et al. 2008). This could reflect a shift to higher trophic level prey items during the breeding period (i.e. kleptoparasitism by frigatebirds), selective feeding of food items to young, age specific fractionation, or some effect of regurgitation of food. Other studies have shown differential provisioning of chicks with higher
quality food, or different trophic level food sources (Hodum & Hobson 2000, Cherel et al. 2008). While seabird studies have not documented changes in $\Delta$N with age, this has been seen in other taxa and could be a viable explanation (Roth & Hobson 2000). Even without different $\Delta$N, by feeding on partially digested food, they may be incorporating $^{15}$N from their parents’ bodies during the regurgitation process, thus explaining higher $^{13}$N values. Direct comparison of adult and chick blood and diets (not taken here) would help resolve this. The lack of any shift in $^{13}$C from adults to chicks was unexpected given that breeding places constraints on seabird foraging distances, but is consistent with the lack of change seen in Seychelles (Catry et al. 2008).

We saw limited evidence of resource partitioning by sex. All 4 species that we examined for sexual partitioning of resources exhibited reverse sexual dimorphism. Of these, we saw differences in $^{15}$N and $^{13}$C in only 1 species, the brown booby. The larger females showed higher $^{15}$N and $^{13}$C, indicating that female brown boobies had a higher trophic level and less pelagic food sources. This is consistent with other evidence of niche partitioning by sex in brown boobies (Weimerskirch et al. 2009b). Likewise, the lack of resource partitioning by sex in red-footed and masked boobies is also consistent with other studies (Weimerskirch et al. 2009a, Young et al. 2010). However, sex differences have been observed in red-footed boobies in other locations (Weimerskirch et al. 2006).

In the 1 species for which we compared data across multiple years, we saw small but significant differences in $^{15}$N for 1 of the 3 years. This may point to small variability in food sources across time even in tropical resources. While it is not always possible to gather samples from all species simultaneously, this suggests caution is needed in interpreting isotopic data from seabirds gathered in different years as evidence of niche partitioning.

**Comparisons to other predators**

We saw clear distinctions among seabirds and other marine predators. There was relatively little overlap between pelagic predators and seabirds on a species by species basis. The primary differences in seabirds and predatory fish was the lower $^{13}$C of seabirds. This difference may reflect the integration of food from the deep scattering layer and reef (both of which would have higher $^{13}$C signatures) into the diets of these fish. This is consistent with other studies reporting that pelagic feeding seabirds have relatively low $^{13}$C values compared to benthic feeding species (Thompson et al. 2000). Although bird and fish predators often forage together, studies of tropical seabird and other predator diets have shown substantial variation among species (Catry et al. 2009). For *Sthenoteuthis* spp., the extremely offshore foraging patterns may explain the more negative values in $^{13}$C observed, and thus the lack of differentiation from seabirds. The difference in $^{15}$N between *Sthenoteuthis* spp. and seabirds may reflect the lower trophic level diet observed in *Sthenoteuthis* spp. (Shchetinnikov 1992).

**CONCLUSIONS**

In general, our study supports the idea that there is significant niche partitioning among tropical seabirds even during non-breeding periods, when many species are utilizing similar open ocean environments. There is overlap in resource use among species during both breeding and non-breeding periods, but when both periods are considered, each species appears to use a unique niche. However, analyses of prey item isotope levels, controlled laboratory experiments of species-specific fractionation rates, and better isotopic maps for the region would help improve the interpretation of such isotopic results.

Tropical seabirds have a more limited suite of foraging techniques than do temperate species, as resources tend to be patchier and more limited in the tropical ocean than in temperate regions. Yet, our results confirm and strengthen results from earlier studies (e.g. Spear et al. 2007) that suggest that niche partitioning is similarly prevalent among seabirds in tropical systems as in temperate systems.

**Acknowledgements.** We thank the National Science Foundation, the National Geographic Society, the Stanford VPUE summer field studies grants, the Stanford Gabilan Graduate Fellowship, the Woods Institute of the Environment, and the Tagging of Pacific Pelagics program for financial or in-kind support. For logistical and material support, we thank the US Fish and Wildlife Service, the Palmyra Atoll National Wildlife Refuge, The Nature Conservancy, and the Palmyra Atoll Research Consortium. For genetic analysis of sex, we thank F. Hailer and E. A. Schreiber at the Smithsonian Institute. For assistance in the field and laboratory, we thank D. Mucciaroni, K. Pollock, J. Svendson, S. Barclay, C. Depkin, A. Briggs, M. deGraff, C. Burniske, and L. Palumbi.

**LITERATURE CITED**

- Ashmole NP (1968) Body size, prey size, and ecological segre-


Cherel Y, Hobson KA, Hassan S (2005a) Isotopic discrimination between food and blood and feathers of captive penguins: implications for dietary studies in the wild. Physiol Biochem Zool 78:106–115


Graham BS (2009) Trophic dynamics and movements of tuna in the tropical Pacific Ocean inferred from stable isotope analyses. PhD dissertation, University of Hawai‘i, Honolulu, HI


Mar Sci 45:52–67

Editorial responsibility: Yves Cherel, Villiers-en-Bois, France

Submitted: May 3, 2010, Accepted: August 2, 2010
Proofs received from author(s): September 23, 2010